

C1 FILE FORMATS

Key Notes

Three common sequence formats

There are several conventions for representing nucleic acid and protein sequences, of which the NBRF/PIR, FASTA and GDE formats are widely used. These formats have limited facilities for comments, which must include a unique identifier code and the sequence accession number.

Files for aligned sequences

Aligned sequences can be represented in NBRF/PIR, FASTA or GDE formats but there are other formats devised especially for multiple sequence alignment, including MSF, PHYLIP and ALN.

Files of structural data

Structural data are maintained as flat files using the PDB format. Such files contain orthogonal atomic co-ordinates together with annotations, comments and experimental details.

Related topics

Annotated sequence databases (C2) Obtaining, viewing and analyzing
Multiple sequence alignment and structural data (I4)
family relationships (F1)

Three common sequence formats

If biological data is to be used by computer programs, it must be presented in a standard format that can be read by computer. It is very common to put data in text files. As the name suggests, these files contain text that can be read by a human being as well as a computer. They are rather like the files used by word-processing packages to hold documents, but there is one important difference: text files hold (almost) only the text and little auxiliary information about formatting (more details below). Here we discuss some standard formats and some more database specific formats are discussed in Topic C2.

Many bioinformatic databases and software applications are designed to work with sequence data, and this requires a standard format for inputting nucleic acid and protein sequence information. Three of the most common sequence formats are NBRF/PIR (National Biomedical Research Foundation/Protein Information Resource), FASTA and GDE. Each of these formats has facilities not only for representing the sequence itself, but also for inserting a unique code to identify the sequence and for making comments which may include for example the name of the sequence, the species from which it was derived, and an accession number for GenBank or another appropriate database (Topic C2). Figure 1 shows the same protein sequence, that of a guinea-pig serotonin receptor, represented in the three sequence formats listed above.

Figure 1a shows the NBRF/PIR format. Note that the first line begins with '>P1;' which specifies a protein sequence. If this was a nucleic acid sequence, it would begin with '>N1;'. The semicolon is followed by a code, in this case '5H1B_CAVPO', which is a unique sequence identifier. Serotonin is also known as 5-hydroxytryptamine, thus 5H1B identifies the protein as serotonin receptor 1B, while CAVPO identifies its source as the guinea-pig (*Cavia porcellus*). There

(a)
 >P1:5H1B_CAVPO
 Guinea pig serotonin receptor accession: 008892
 MGNPEASCTP PAVLGSQTGL PHANVSAPPN NCSAPSHIYQ DSIALPWKVL LVVLLALITL
 ATTLNSAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL VMPISTMYTV TGRWTLGQAL
 CDFWLSSDIT CCTASIMHLC VIALDRYWAI TDAVGYSAGR TPRRAAGMIA LVWVFSICIS
 LPPFFWRQAK AEEVLDCLV NTDHVLTVY STGGAFYLP LLLIALYGRI YVEARSRLK
 QTPNKTGKRL TRAQLITDSP GSTSSVTSIN SRAPEVPCDS GSPVYVQVK VRVSDALLEK
 KKLMAARERK ATKTLGVILG AFIVCWLPFF IISLVMPICK DACWFHMAIF DFFTWLGYLN
 SLINPIIYTM SNEDFKQAFH KLIRFKCTT
 *

(b)
 > 5H1B_CAVPO 008892|guinea pig serotonin receptor
 MGNPEASCTP PAVLGSQTGL PHANVSAPPN NCSAPSHIYQ DSIALPWKVL LVVLLALITL
 ATTLNSAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL VMPISTMYTV TGRWTLGQAL
 CDFWLSSDIT CCTASIMHLC VIALDRYWAI TDAVGYSAGR TPRRAAGMIA LVWVFSICIS
 LPPFFWRQAK AEEVLDCLV NTDHVLTVY STGGAFYLP LLLIALYGRI YVEARSRLK
 QTPNKTGKRL TRAQLITDSP GSTSSVTSIN SRAPEVPCDS GSPVYVQVK VRVSDALLEK
 KKLMAARERK ATKTLGVILG AFIVCWLPFF IISLVMPICK DACWFHMAIF DFFTWLGYLN
 SLINPIIYTM SNEDFKQAFH KLIRFKCTT

(c)
 5H1B_CAVPO 008892|guinea pig serotonin receptor
 MGNPEASCTP PAVLGSQTGL PHANVSAPPN NCSAPSHIYQ DSIALPWKVL LVVLLALITL
 ATTLNSAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL VMPISTMYTV TGRWTLGQAL
 CDFWLSSDIT CCTASIMHLC VIALDRYWAI TDAVGYSAGR TPRRAAGMIA LVWVFSICIS
 LPPFFWRQAK AEEVLDCLV NTDHVLTVY STGGAFYLP LLLIALYGRI YVEARSRLK
 QTPNKTGKRL TRAQLITDSP GSTSSVTSIN SRAPEVPCDS GSPVYVQVK VRVSDALLEK
 KKLMAARERK ATKTLGVILG AFIVCWLPFF IISLVMPICK DACWFHMAIF DFFTWLGYLN
 SLINPIIYTM SNEDFKQAFH KLIRFKCTT

Fig. 1. The sequence of a guinea-pig serotonin receptor in (a) NBRF/PIR format; (b) FASTA format; and (c) GDE format.

follows a **comment line**, and the rules allow this line to be of more or less any length so it can either be empty or far too wide to fit on a printed page. Then the sequence itself follows and is terminated by an asterisk (*). It is conventional to give files in this format the extension '.pir' or '.seq'.

Figure 1b shows the **FASTA format**. The first line begins with '>' but there is no designation of protein or nucleic acid sequence. The code is entered next and this is followed (on the same line) by comments, although it is conventional to delimit the comments with a '|' symbol. As with the NBRF/PIR format there is no limit to the length of the first line. One point to note about FASTA files is that they allow lower-case letters for the amino acids. Files in this format commonly have the extension '.fasta'.

Figure 1c shows the **GDE format**. This is essentially the same as the FASTA format, but the '>' symbol in the first line is replaced by '%'. Files in this format have the extension '.gde'.

All three file formats *ignore spaces and carriage returns*. This allows sequences to be typed out in a manner that is convenient for the user. In Fig. 1, for example, a space has been inserted every 10 amino acid residues and a carriage return after every 60, making it much easier to manually count the residues and identify amino acids at specific positions in the sequence. Note, however, that most standard word-processing software packages do not ignore blank spaces. For some purposes, it may be desirable or necessary to construct files from unpublished and preliminary sequence data, and if programs such as Microsoft Word or Corel WordPerfect are used, the results can be unpredictable. If using Word, use text only mode with a non-proportional font or, preferably, use a simple text editor such as Notepad.

Files
sequ

Files
struc

To illustrate this point, consider the creation of the following very simple NBRF/PIR file:

```
>P1;MY_CODE
my cat
MYCATSATINMYLAP*
```

Despite the fact that this protein is clearly fictitious, the format is perfectly correct and it should be possible to search for the peptide sequence in other proteins. However, by typing this sequence into Microsoft Word and saving it as a Word document (cat.doc), the file proves to be over 19 thousand bytes in length, and therefore obviously contains much more than the simple text. By saving the file as *text with line breaks* (cat.txt), the file size is reduced to 39 bytes, which seems more reasonable. However, inspection of the contents of cat.txt reveals two extra characters at the end of each line and another at the very end of the file. It is therefore best to avoid word processors and use text editors for the preparation of sequences. If a word processor is used, the file should be saved as text and sent by ftp as ASCII (Topic O4), and a text editor should then be used on the computer where the sequence analysis is carried out, to check the integrity of the file. Another point to bear in mind is that the first line of a FASTA file and the second line of an NBRF/PIR file might be extremely long and it is essential not to cut it up by inserting carriage returns, otherwise the comments might be read as part of the sequence.

Files for aligned sequences

The output from sequence-alignment programs can be in any one of a number of formats. All three formats discussed above are suitable for dealing with aligned sequences but there are several formats designed specifically for alignment output. Figure 2 shows partial results from the alignment of five serotonin receptor sequences, including the guinea-pig 5H1B receptor. In order to achieve the alignments, gaps must be introduced (Topic E3) and these are represented either by hyphens or dots. Multiple sequence format (MSF) is used by several software tools. PHYLIP (phylogenetic inference package) is the output format of the software of that name and CLUSTALW/X (F1) has its own ALN format. Multiple sequence alignment is discussed in more detail in Topic F1.

Files of structural data

The raw materials for bioinformatic studies on macromolecular structures are PDB files. These are text files using a format devised by the Protein Data Bank (Topic C4). Such files contain orthogonal atomic co-ordinates together with annotations, comments and experimental details. Examples of parts of such files are shown in Fig. 3. The most important aspect of PDB files is that the 'ATOM' lines are laid out in columns of characters *not* columns of words. Compare the first ATOM lines from Fig. 3a, b and c (only the left-hand parts of each line are displayed here):

ATOM	1	N	VAL	16	29.582	19.112	38.968
ATOM	1	N	ILE E	16	-9.947	23.613	20.817
ATOM	1	N	ALA	1	14.702	-10.824	3.425

The last three columns show the orthogonal co-ordinates (x, y, z) of the atom and they are deduced by counting the positions including the spaces, *not* by counting the words. This is because the x co-ordinate is the sixth word in the first and third cases but the seventh word in the second case, because a new

(a)

MSF: 435 Type: P Check: 2299 ..

Name: 5H1A_MOUSE oo Len: 435 Check: 7521 Weight: 0.166
 Name: 5H1A_RAT oo Len: 435 Check: 8470 Weight: 0.250
 Name: 5H1A_HUMAN oo Len: 435 Check: 8517 Weight: 0.166
 Name: 5H1B_CAVPO oo Len: 435 Check: 829 Weight: 0.222
 Name: 5H1B_CRIGR oo Len: 435 Check: 6962 Weight: 0.100

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5H1A_MOUSE      MD.....MF SLGQGNNTTT SLEPFG.... ..TGGNDTGL SNVTFSYQVI
5H1A_RAT        MD.....VF SFGQGNNTTA SQEPFG.... ..TGGNVTST SDVTFSYQVI
5H1A_HUMAN      MD.....VL SPGQGNNTTS PPAPFE.... ..TGGNTTGI SDVTFSYQVI
5H1B_CAVPO      MGNPEASCTP PAVLGSQTGL PHANVSAPPN NCSAPSHIYQ DSIALPWKVL
5H1B_CRIGR      MEEQGIQCAP PPPAASQTGV PLVNLS...H NCSAESHYQ DSIALPWKVL

5H1A_MOUSE      TSLLLGTLIF CAVLGNACVV AAIALERSLQ NVANYLIGSL AVTDLMSVSL
5H1A_RAT        TSLLLGTLIF CAVLGNACVV AAIALERSLQ NVANYLIGSL AVTDLMSVSL
5H1A_HUMAN      TSLLLGTLIF CAVLGNACVV AAIALERSLQ NVANYLIGSL AVTDLMSVSL
5H1B_CAVPO      LVLLALITL ATTLNFAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL
5H1B_CRIGR      LVLLALITL ATTLNFAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL

```

{ rest of file omitted }

(b)

```

      5      435
5H1A_MOUSE MD-----MF SLGQGNNTTT SLEPFG---- --TGGNDTGL SNVTFSYQVI
5H1A_RAT   MD-----VF SFGQGNNTTA SQEPFG---- --TGGNVTST SDVTFSYQVI
5H1A_HUMAN MD-----VL SPGQGNNTTS PPAPFE---- --TGGNTTGI SDVTFSYQVI
5H1B_CAVPO MGNPEASCTP PAVLGSQTGL PHANVSAPPN NCSAPSHIYQ DSIALPWKVL
5H1B_CRIGR MEEQGIQCAP PPPAASQTGV PLVNLS---H NCSAESHYQ DSIALPWKVL

      TSLLLGTLIF CAVLGNACVV AAIALERSLQ NVANYLIGSL AVTDLMSVSL
      TSLLLGTLIF CAVLGNACVV AAIALERSLQ NVANYLIGSL AVTDLMSVSL
      TSLLLGTLIF CAVLGNACVV AAIALERSLQ NVANYLIGSL AVTDLMSVSL
      LVLLALITL ATTLNFAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL
      LVLLALITL ATTLNFAFVI ATVYRTRKLH TPANYLIASL AFTDLLVSIL

```

{ rest of file omitted }

(c)

>P1; 5H1A_MOUSE

```

MD-----MFSLGQGNNTTTTSLPFG-----TGGNDTGLSNVTFSYQVITSLLLGTLIF
CAVLGNACVVAAIALERSLQNVANYLIGSLAVTDLMSVSLVLPMAALYQVLNKWTLGQVT
CDLFIALDVLCTSSILHLCAIALDRYWAITDPIDYVNRTPRRAAALISLTWLIGFLIS
IPPMLGWRAPEDRSNPNECTISKDHG-YTIYSTFGAFYIPLLLMLVLYGRIFRAARFRIR
KTVRKVEKKGAGTSPGTSSAPPPKSLNGQPGSGDCRRSAENRAVGTPCANGAVRQGEDD
ATLEVIEVHRVGNSKGDLPLPSESGATSYPACLERKNERTAEAKRKMALARERKTVKTL
GIIMGTFILCWLPPFIVLVLPPCESSCHMPPELLGAIINWLGYSNLLNPVIYAYFNKDF
QNAFKKIICKKPCR-

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>P1; 5H1A_RAT

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MD-----VPSFGQGNNTTASQEPFG-----TGGNVTSTSDVTFSYQVITSLLLGTLIF
CAVLGNACVVAAIALERSLQNVANYLIGSLAVTDLMSVSLVLPMAALYQVLNKWTLGQVT
CDLFIALDVLCTSSILHLCAIALDRYWAITDPIDYVNRTPRRAAALISLTWLIGFLIS
IPPMLGWRTPEDRSNPDCTISKDHG-YTIYSTFGAFYIPLLLMLVLYGRIFRAARFRIR
KTVRKVEKKGAGTSLGTSSAPPPKSLNGQPGSGDWRRCAENRAVGTPTNGAVRQGDDE
ATLEVIEVHRVGNSKEHLPLPSESGSNSYAPACLERKNERNABAKRKMALARERKTVKTL
GIIMGTFILCWLPPFIVLVLPPCESSCHMPALLGAIINWLGYSNLLNPVIYAYFNKDF
QNAFKKIICKKPCR

```

{ rest of file omitted }

Fig. 2. Partial results from the alignment of five proteins with CLUSTALW (Topic F1). The formats shown are (a) MSF output; (b) PHYLIP output; and (c) NBRF/PIR output.

(a) Trypsin

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HEADER    HYDROLASE (SERINE PROTEINASE)          13-APR-88  180T  180T  3
COMPND    TRYPSIN (/SGT$) (E.C.3.4.21.4)         180T  4
SOURCE    (STREPTOMYCES SGRISEUS, STRAIN K1)     180T  5
AUTHOR    R.J.READ,M.N.G.JAMES                   180T  6
REVDAT    1 16-JUL-88 180T 0                     180T  7
JRNL      There follow the literature references
REMARK    1 There follow several remarks of which only 1 is shown here 180T 21
REMARK    2 RESOLUTION. 1.7 ANGSTROMS.           180T 72
           The sequence (only 2 lines shown) follows
SEQRES    1 223 VAL VAL GLY GLY THR ARG ALA ALA GLN GLY GLU PHE PRO 180T 92
SEQRES    2 223 PHE MET VAL ARG LEU SER MET GLY CYS GLY GLY ALA LEU 180T 93
FTNOTE    1 There follow several footnotes       180T 110
MET        CA 246 1 CALCIUM ++ ION              180T 151
FORMUL    2 CA CA1 ++                           180T 152
FORMUL    3 HOH *192(H2 O) The last 3 lines describe hetero atoms 180T 153
           there follow several lines of secondary structure assignment
           of which only the first is shown here
HELIX      1 A ALA 56 CYS 58 5                   180T 154
           There follow 7 lines describing the orthogonal coordinate system
ATOM       1 N VAL 16 29.582 19.112 38.968 1.00 12.94 180T 199
ATOM       2 CA VAL 16 30.031 20.461 38.668 1.00 15.43 180T
           ....the bulk of the file
ATOM       1618 CD1 LEU 245 2.571 16.977 47.866 1.00 40.15 180T1816
ATOM       1619 CD2 LEU 245 4.758 18.112 48.337 1.00 44.30 180T1817
ATOM       1620 OXT LEU 245 1.660 16.559 52.387 1.00 59.60 180T1818
TER        1621 LEU 245                          180T1819
HETATM     1622 CA CA 246 14.219 32.828 30.463 1.00 13.21 180T1820
HETATM     1623 O HOH 1 22.919 19.524 42.538 1.00 8.79 180T1821
           ....the remaining water molecules up to ...
HETATM     1814 O HOH 192 -3.192 30.325 46.346 0.68 57.70 180T2012
CONECT     72 70 941                             180T2013
           ....the connectivity data
MASTER    79 41 1 5 14 15 1 6 1813 1 30 18 180T2043
END                                                180T2044

```

(b) Complex of a proteinase ("E") with a polypeptide inhibitor ("I")

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HEADER    COMPLEX (SERINE PROTEINASE-INHIBITOR)  21-JAN-83  38GB  38GBE 1
COMPND    PROTEINASE B FROM STREPTOMYCES GRISEUS (/SGPBS) 38GB 4
COMPND    2 (E.C. NUMBER NOT ASSIGNED) COMPLEX WITH THIRD DOMAIN OF THE 38GB 5
COMPND    3 TURKEY OVOMUCOID INHIBITOR (/OMTKY3$) 38GB 6
SOURCE    (STREPTOMYCES SGRISEUS, STRAIN K1) AND TURKEY (MELEAGRIS 38GB 7
SOURCE    2 GALLOPAVO) 38GB 8
AUTHOR    R.J.READ,M.FUJINAGA,A.R.SIELECKI,M.N.G.JAMES 38GB 9
           There follow many lines of remarks and details of lit. references
           One such remark is important
REMARK    2 RESOLUTION. 1.8 ANGSTROMS.           38GB 73
           Start of sequence ...
SEQRES    1 E 185 ILE SER GLY GLY ASP ALA ILE TYR SER SER THR GLY ARG 38GB 92
           Start of ATOM entries for "chain E" ...
ATOM       1 N ILE E 16 -9.947 23.613 20.817 1.00 16.42 38GB 156
           ...end of "chain E" and start of "chain I"
ATOM       1310 OXT TYR E 242 -10.317 35.858 21.204 1.00 29.02 38GB1465
TER        1311 TYR E 242                        38GB1466
ATOM       1350 N ASP I 7 25.100 14.110 33.198 1.00 41.61 38GB1467
ATOM       1351 CA ASP I 7 25.863 15.369 33.122 1.00 40.76 38GB1468
           ... to the end of the file

```

(c) Intestinal fatty acid binding protein

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HEADER    FATTY ACID-BINDING                     20-FEB-98  1A57
TITLE     THE THREE-DIMENSIONAL STRUCTURE OF A HELIX-LESS VARIANT OF
TITLE     2 INTESTINAL FATTY ACID BINDING PROTEIN, NMR, 20 STRUCTURES
COMPND    MOL_ID: 1;
COMPND    2 MOLECULE: INTESTINAL FATTY ACID-BINDING PROTEIN;
           Many lines of remarks including the authors but 3 are included:
EXPDTA    NMR, 20 STRUCTURES
AUTHOR    R.A.STEELE,D.A.EMMERT,J.KAO,M.E.HODSDON,C.FRIEDEN,
AUTHOR    2 D.P.CISTOLA
REMARK    Compare the following line with its counterparts in (a) and (b)
REMARK    2 RESOLUTION. NOT APPLICABLE.
           Start of the sequence
SEQRES    1 116 ALA PHE ASP GLY THR TRP LYS VAL ASP ARG ASN GLU ASN
           Start of the secondary structure assignment
SHEET     1 A 5 GLY 4 LYS 7 0
MODEL     1 start of the atomic coordinates for "model 1"
ATOM       1 N ALA 1 14.702 -10.824 3.425 1.00 0.00 N
ATOM       2 CA ALA 1 13.562 -10.618 2.552 1.00 0.00 C
ATOM       3 C ALA 1 12.273 -10.914 3.355 1.00 0.00 C
           ... up to the end of that and start of "model 2"
ATOM       2132 OG GLU 116 15.846 0.773 9.258 1.00 0.00 Q
TER        2133 GLU 116
ENDMDL
MODEL     2
ATOM       2134 N ALA 1 14.997 -8.697 2.368 1.00 0.00 N
ATOM       2135 CA ALA 1 14.960 -9.149 3.746 1.00 0.00 C
           ... up to the end (there is a total of 20 such models).

```

Fig. 3. Parts of three PDB files showing (a) and (b) X-ray crystallographic data; and (c) NMR data. Comments not in the original files but added here for clarity are shown in *italic*.

word, the chain identifier (E), has been inserted. Other points that emerge from Fig. 3 are: the atoms are numbered consecutively; amino acids are represented by three letters; the ends of the lines may or may not contain line numbers; NMR files do not have a *resolution* REMARK; and NMR files typically contain several models corresponding to different conformations.